Data pre-processing before running CEIIFi

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January 12, 2023

CEIIFi is a python package that models DNA methylation calls from references with non-negative least squares and estimate the fraction of cell types in a DNA mixture.

$$argmin_x||Ax-b||_2^2$$

with the constraint $x \ge 0$, $\sum x = 1$, A denotes an $m \times n$ matrix.

Dependencies

- python2, [Recommend] Building the environment with virtualenv
 - pandas
 - numpy
 - scipy
- CELLFi package
 - Click for reference .bed files and scripts, or contact Dr. Dennis Montoya for Github access

Differentially methylated regions (DMRs)

Here we demonstrate the CEllFi deconvolution with blood cells' whole genome bisulfite sequencing (WGBS) dataset **GSE186458**. Thirty six blood cell WGBS libraries are included for references:

- GSM5652277_Blood-T-CD3-Z000000TV
- GSM5652278_Blood-T-CD3-Z000000UP
- GSM5652279_Blood-T-CD4-Z000000TT
- GSM5652280_Blood-T-CD4-Z000000U7
- GSM5652281_Blood-T-CD4-Z000000UM

- GSM5652282_Blood-T-CD8-Z000000TR
- GSM5652283_Blood-T-CD8-Z000000U5
- GSM5652284_Blood-T-CD8-Z000000UK
- \bullet GSM5652285_Blood-T-CenMem-CD4-Z00000417
- \bullet GSM5652286_Blood-T-CenMem-CD4-Z0000041D
- $\bullet \ \ GSM5652287_Blood-T-CenMem-CD4-Z0000041N$
- GSM5652288_Blood-T-Eff-CD8-Z00000419
- \bullet GSM5652289_Blood-T-Eff-CD8-Z0000041F
- \bullet GSM5652290_Blood-T-Eff-CD8-Z0000041Q
- GSM5652291_Blood-T-EffMem-CD4-Z00000416
- \bullet GSM5652292_Blood-T-EffMem-CD4-Z0000041C
- \bullet GSM5652293_Blood-T-EffMem-CD4-Z0000041M
- \bullet GSM5652294_Blood-T-EffMem-CD8-Z0000041A
- \bullet GSM5652295_Blood-T-EffMem-CD8-Z0000041G
- GSM5652296_Blood-T-Naive-CD4-Z0000041E
- \bullet GSM5652297_Blood-T-Naive-CD8-Z0000041B
- GSM5652298_Blood-T-Naive-CD8-Z0000041H
- \bullet GSM5652299_Blood-NK-Z000000TM
- \bullet GSM5652300_Blood-NK-Z000000U1
- \bullet GSM5652301_Blood-NK-Z000000UF
- GSM5652302_Blood-Monocytes-Z000000TP
- GSM5652303_Blood-Monocytes-Z000000U3
- GSM5652304_Blood-Monocytes-Z000000UH
- $\bullet \ \ GSM5652313_Blood\text{-}Granulocytes\text{-}Z0000000TZ$
- $\bullet \ \ GSM5652314_Blood\text{-}Granulocytes\text{-}Z000000UD$
- \bullet GSM5652315_Blood-Granulocytes-Z000000UT
- GSM5652316_Blood-B-Z000000TX

- GSM5652317_Blood-B-Z000000UB
- GSM5652318_Blood-B-Z000000UR
- GSM5652319_Blood-B-Mem-Z0000041J
- GSM5652320_Blood-B-Mem-Z0000041K

We aim to build references for B cell, monocyte, granulocyte, nature killer cell (NK cell), and T cell. To achieve this, we first identify cell-type specific hypo-DMRs with the *metilene* software:

```
metilene -M 500 -m 10 -d 0.3 -t 2 -v 0.7 --mode 1 --mtc 2
--groupA 'Bcell_' --groupB 'nonBcell_' Bcell_metilene_input.tsv
> Bcell_DMR.tsv
```

The above example demonstrates that we demand DMRs with at least 500bp in size with minimum 10 CpG sites that show differential methylation level >=30% between B cell and the other cell types. Further filtering could be applied to acquire hypo-DMRs based on the delta methylation level in column 4 of the output Bcell_DMR.tsv file. We further characterize naïve T cell's hypo-DMRs from other T cell.

To verify the DMRs, we performed supervised hierarchical clustering:

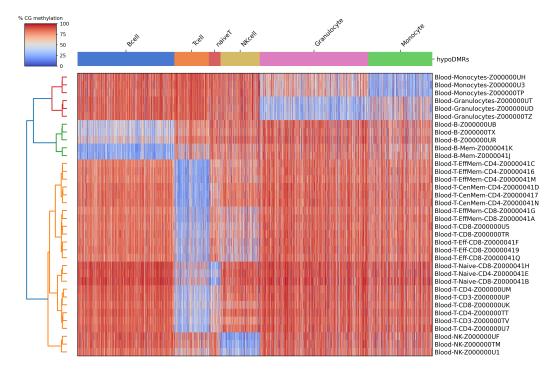


Figure 1: All cell-specific DMRs (n= 110,276)

CpG in these DMRs could be used as references for CEllFi deconvolution. However, it could take hours for CEllFi to construct reference matrix, and, when applied to targeted-bisulfite sequencing data (TBS), the joint matrix of reference and sample would have *nan* in majority, which is not cost-effective. Subset is therefore recommended. The following cluster heatmap shows that the only 60 DMRs, subset from the COVID19 TBS dataset, still are able to distinguish the 6 cell types:

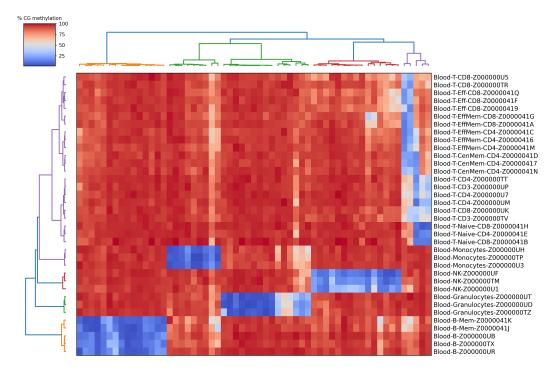


Figure 2: TBS subset cell-specific DMRs (n= 60)

After subsetting the CpG, all reference .bed files are moved to ./ref/ directory.

Validation

Three .conf files need to be ready before running CEIIFi:

- ullet cellfi_reference.conf, binary classification pointing the reference .bed to the cell type
- cellfi_sample.conf, sample .bed to be deconvoluted
- cellfi_group.conf, cell types to be included in the analysis

Make sure the reference and sample .bed files are in the correct directory, and run:

```
FILE Bcell Tcell NKcell Monocyte Granulocyte NaiveT

./ref/GSM5652277_Blood-T-CD3-Z000000TV_all_celltype-specific_hypoDMR_mCG.bed 0 1 0 0 0

./ref/GSM5652278_Blood-T-CD3-Z000000TV_all_celltype-specific_hypoDMR_mCG.bed 0 1 0 0 0

./ref/GSM5652279_Blood-T-CD4-Z000000TT_all_celltype-specific_hypoDMR_mCG.bed 0 1 0 0 0

./ref/GSM5652280_Blood-T-CD4-Z0000000TY_all_celltype-specific_hypoDMR_mCG.bed 0 1 0 0 0

./ref/GSM5652281_Blood-T-CD4-Z000000UM_all_celltype-specific_hypoDMR_mCG.bed 0 1 0 0 0

./ref/GSM5652282_Blood-T-CD8-Z000000UTR_all_celltype-specific_hypoDMR_mCG.bed 0 1 0 0 0

./ref/GSM5652282_Blood-T-CD8-Z000000UTR_all_celltype-specific_hypoDMR_mCG.bed 0 1 0 0 0
```

Figure 3: Example of cellfi_reference.conf

```
FILE
./samples/natural_killer_cell_C002CTA1bs_hg38.bed
```

Figure 4: Example of cellfi_sample.conf

```
FILE
./samples/natural_killer_cell_C002CTA1bs_hg38.bed
```

Figure 5: Example of cellfi_group.conf

```
python ./scripts/bs_decon_pipe.py -pipe_o ./ -estimate_refs \\
Bcell,Granulocyte,Monocyte,NKcell,NaiveT,Tcell \\
-detect_delta 0.30 -select_num_hypo 15 -select_ttest_p 0.1 \\
-select_anova_p 0.1 -select_sam_cpg 5 -select_cell_delta 0.3
```

```
Samples Bcell Granulocyte Monocyte NKcell NaiveT Residual Tcell
CD4_CD45_RA_naive_5277_covered 0.0 0.003725486266468322 0.0 0.0521421761117374 0.9398169890433812 0.37926152288397524 0.0
```

Figure 6: Example of cellfi_coeff_meth.txt

Noted that parameters could be adjusted if needed. Several intermediate .txt files will be generated, and the cellfi_coeff_meth.txt is the final output that shows the fraction of denoted cell types in the query sample.

In order to validate the CEIIFi deconvolution, we took the following WGBS data (in the ./samples/ directory):

- CD4_CD45_RA_naïve_5277, pellegrini lab
- CD4_CD45_RO_memory_5277, pellegrini lab
- classical_monocyte_C000S5A1bs, blueprint
- mature_neutrophil_C000S5A2bs, blueprint
- natural_killer_cell_C002CTA1bs, blueprint
- in silico synthetic mixture of 30% B cell and 70% Monocyte
- in silico synthetic mixture of 70% B cell and 30% Monocyte

Figure 7 bellow shows that all WGBS are with > 90% composition of the indicated cell types.

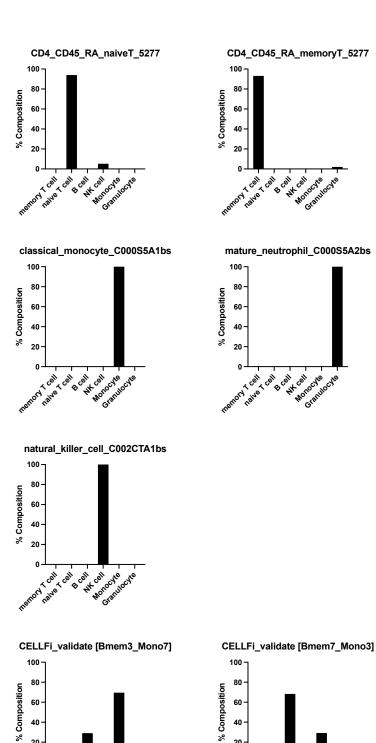


Figure 7: CEIIFi validation

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