HOMOLOGY MODELLING



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Homology Modelling

What is it and why do we need it?

principles of modelling, applications available

Using Swiss-Model

- preparation, workspace tools, automated-mode
- refining with manually-adjusted alignments

Model Assessment

- using SwissModel tools to check model quality
- what to look for, what we expect...

What is Homology Modelling?

- Given an unknown protein, make an informed guess on its 3D structure based on its sequence:
 - Search structure databases for *homologous* sequences
 - Transfer coordinates of known protein onto unknown

MQEQLTDFSKVETNLISW-QGSLETVEQMEPWAGSDANSQTEAY



= Identity

. | = Homology

Why Modelling is Necessary

Current structure determination methods:

- NMR conc. protein solution; limited size + resolution
- XRay Crystallography protein crystals, high resolution
- Expensive, slow, difficult (especially membrane proteins!)

Can't keep up with growth rate of sequence databases:

- PDB: 40132 structures (14/11/2006)
- pairsdb: 4000000+ sequences (11/2006)

Current (Free) Servers & Software

- SWISSMODEL (www) http://swissmodel.expasy.org/SWISS-MODEL.html
- 3D-Jigsaw (www) http://www.bmm.icnet.uk/servers/3djigsaw/
- ESyPred3D (www) http://www.fundp.ac.be/sciences/biologie/urbm/bioinfo/esypred/
- WHATIF (www) http://swift.cmbi.kun.nl/WIWWWI
- CPH Models 2.0 (www) http://www.cbs.dtu.dk/services/CPHmodels
- MODELLER 8v2 (standalone for windows, mac, linux; also web submission) http://salilab.org/modeller http://alto.compbio.ucsf.edu/modweb-cgi/main.cgi

Assumptions & Principles

- Increase in sequence identity correlates with increase in structural similarity
- RMSD of core α-carbon coordinates for two homologous proteins sharing 50% identity expected at ~1Å (GLY 3.5Å, helix 5Å diameter)
- Theoretical models are low resolution, and depend on quality of input alignment!

The SwissModel Workspace

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Preparation

Automated mode:

 Select a template structure with high % identity and average resolution, or let SwissModel choose best one.

Alignment mode:

- Obtain multiple alignment between possible templates and your query sequence.
- Check alignment are key motifs correctly aligned? Manually adjust as necessary, using Jalview.
- Upload alignment, select template, and submit.

Submit a Model Request

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Submit a Model Request



Hidden Modelling Steps

Automated mode, multiple templates:

- Superimposition and optimisation of corresponding α-carbon pairs for all template structures
- Alignment of all residues of template structures: acceptable alpha-carbon atoms located within 3Å radius of mean

All modes:

- Framework construction of query protein structure, using coordinates of template structure, or mean coordinate positions if multiple templates.
- Building unconserved loop regions / insertions, closing gaps in backbone - uses penta-peptide fragment library derived from all PDB entries of resolution 2Å or better (note: this can fail badly!)

Output

- Within a few minutes of submission, your results are returned to the Workspace.
- Output Page:

 Your model in pdb format (plus simple viewer)
 Query to template alignment
 Simple assessment graphs
 Logging data of the modelling process
- Save the model in DeepView Project format, then open in DeepView, and colour by B-factor.



Multiple Sequence Alignment

- More information than pairwise alignment
- Shows conserved regions in protein family
 - Critical secondary structure elements
- Shows strongly conserved residues and motifs
 - Structural importance
 - Functional importance

Alignment Adjustment

Look at alignment used to select template

- Check alignment against 3D structure (Jalview)
- Secondary structure elements preserved?
- Motifs and key residues aligned?
- Are gaps in acceptable locations (ie: loops)?
- Try to improve alignment manually ...
- Or use different alignment application



The SwissModel Repository

- Contains theoretical models (996876 on 05/09/2006)
- Searchable by UniProt / Trembl accession number
- Fully automated procedure models may be bad!
- It can be found here: http://swissmodel.expasy.org/repository
- ModBase also holds threshold-validated theoretical models [4064704 structures currently]: http://alto.compbio.ucsf.edu/modbase-cgi/index.cgi

Model Assessment

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Model Evaluation

- Look at model in DeepView: is it globular and compact, are there odd unstructured loops?
- Colour by B-factor and note bad areas in loops, or in important secondary elements?
- WhatCheck always lots of errors! Look for packing, inside-outside, torsion angle errors.
- Check Solvation Profile how well packed is the model? Are there regions which are too loose, and therefore solvent-exposed?

Torsion Angles



Ramachandran Plot



Experimentally Resolved Model (NMR, XRay):

 Rare for residues other than Glycine & Alanine to be in disallowed regions

Homology Model:

- Bad residues common
- May have to remodel small sections and loops
- Large residues in bad regions worth further investigation ...

Solvation Profile



- Long stretches above zero probably loops
- Most-negative regions well buried in core
- Functional residues above zero!
- Majority of residues must be below zero
- Compare your model to pdb template!

Further details

DeepView tutorial

http://au.expasy.org/spdbv/text/tutorial.htm

Advanced tutorial on SwissModel & DeepView

http://www.usm.maine.edu/~rhodes/SPVTut/text/HGHomMod.html

Suggested Reading

Swiss-Model's advice:

http://www.expasy.org/swissmod/SM_ModelAccuracy.html

A good interactive tutorial on Protein Modelling:

http://www.ncbi.nlm.nih.gov/Class/minicourses/x1a.html

References

Schwede T, Kopp J, Guex N, and Peitsch MC (2003) SWISS-MODEL: an automated protein homology-modeling server. *Nucleic Acids Research 31:3381-3385.*

Guex, N. and Peitsch, M. C. (1997) SWISS-MODEL and the Swiss-PdbViewer: An environment for comparative protein modelling. *Electrophoresis* 18:2714-2723.

Peitsch, M. C. (1995) Protein modeling by Email. *Bio/ Technology* 13:658-660.

R.W.W.Hooft, G.Vriend, C.Sander and E.E.Abola, Errors in protein structures. *Nature 381, 272 (1996).*

Ramachandran GN, Ramakrishnan C, Sasisekharan V (1963). J Mol Biol 7:95-99.

- Uniprot entry Q11CR8_MESSB is described as a "Fructosebisphosphate aldolase class 1, from Mezorhizobium sp. (strain BNC1)"
- It belongs to Pfam <u>PF00274</u>, the Fructose-1,6-Bisphosphate Aldolases (Class 1); this family contains 24 PDB structures.
- When we run BlastP on Q11CR8_MESSB against the PDB, we get hits to all these structures; %ids range from 48 to 54%, which is reassuringly high.
- Let's try SwissModel in Automated mode:
- <u>http://swissmodel.expasy.org/workspace</u>

- Uniprot entry Q07159 is another Class I FBP Aldolase from Staphylococcus carnosus. It is unusual, however – most bacterial FBP Aldolases belong to Class II (PF00596)
- When we run a Blast against the PDB, again we get hits to all twentyfour PDB structures, but identities range from 25 to 32% – not good.
- Try SwissModel in Automated mode...
- Oh dear, only part of our sequence has been modelled automated mode has failed this time. Pfam has a <u>family alignment</u>, so let's use it as input to Alignment mode.

Now let's assess our two complete models:

- Go to [Tools] >> Structure Assessment, and upload each model in turn:
 - Add Whatcheck and Procheck to the selection, and add the resolution (check result file for automated mode, or check PDB website for alignment mode)
- Also, go to the <u>SolvX Server</u> and input our models. It should give us (at least) two graphs per input – the first will be our model, the other(s) our template(s).

What validation checks are the important ones?

- Whatcheck: Accessibility-related (2.6), 3D-Database (2.8), Geometric (2.5.4 2.5.12 2.5.13 2.5.14) Checks. Make a note of bad residues and regions.
- Procheck: Residues Report (G-factors) and Ramachandran Plot. Again, note bad residues and regions.
- SolvX: Compare your model with the template(s). Where are the worst (most solvent accessible) regions? Note them down.
- How does your alignment model for ALF1_STACA compare with your neighbour's model?